UNITED STATES DISTRICT COURT SOUTHERN DISTRICT OF NEW YORK

ADRIANA AVILES, Individually and as Parent and Natural Guardian of N.A., N.A. and A.A., STEPHANIE DENARO, Individually and as Parent and Natural Guardian of D.D. and H.D., CHRISTINE KALIKAZAROS, Individually and as Parent and Natural Guardian of Y.K., GAETANO LA MAZZA, Individually and as Parent and Natural Guardian of R.L., CRYSTAL LIA, Individually and as Parent and Natural Guardian of F.L., and CHILDREN'S HEALTH DEFENSE,	
Plaintiffs,	DECLARATION OF SIN HANG LEE, MD
Against	Civil No.: 1:20-cv-09829-PGG
BILL de BLASIO , in his Official Capacity as Mayor of the City of New York, DR. DAVID CHOKSHI , in his Official Capacity of Health Commissioner of the City of New York, NEW YORK CITY DEPARTMENT OF EDUCATION, RICHARD A. CARRANZA , in his Official Capacity as Chancellor of the New York City Department of Education and THE CITY OF NEW YORK ,	
Defendants.	

I, Sin Hang Lee, MD declare as follows:

1. I am a pathologist and research scientist based in Connecticut who has been

developing DNA sequencing-based molecular tests to diagnose infectious diseases that are

difficult to diagnose, such as Lyme disease and human papilloma virus (HPV) infection.

2. I have trained and taught in some of the world's most prestigious institutions and

has published scores of scientific articles in peer-reviewed journals.

3. Exhibit 1 is a true and accurate copy of my current CV.

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 2 of 23

4. It is my professional opinion that the need for accurate coronavirus testing is imperative. It is especially critical in nursing homes and institutions caring for elderly patients, so that false-positive patients are not housed with true-positive patients. It's also essential to ensure that staff in direct contact with highly susceptible patients be infection-free.

5. I have been working diligently to overcome the roadblocks in coronavirus testing.

6. As early as March 2020, I wrote to the WHO and to Dr. Anthony Fauci at the National Institute of Allergies and Infectious Diseases of the National Institutes of Health (NIH) to explain why the current tests to detect SARS-CoV-2 RNA are generating false positives and negatives. The letter explained that a two-phased test with DNA sequencing would "guarantee no-false positive results" based on my research and published work from Japan. A true and correct copy of that letter is attached as Exhibit 2. The method I used was subsequently published in a peer-reviewed international journal based in Japan.

7. It is my opinion, one supported by the FDA, that questionable or false RT-qPCR test results can be investigated and resolved by Sanger sequencing, the testing method I developed.

8. Given the specificity of my letter, with its DNA sequencing electropherograms and its significance for preventing disease spread, it is almost unbelievable that as of the date of this declaration, I have received no response from either the WHO or NIH.

9. In April 2020, I reached out to the Connecticut Department of Public Health to receive patient samples for further validation testing. I re-tested 20 reference patient samples supplied by the Connecticut Department of Public Health on April 30, 2020, and found that 3 of 10 reference samples initially classified as positive for SARS-CoV-2 by RT-qPCR test were false positives and 2 of 10 reference samples initially classified as negative were found to

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 3 of 23

contain SARS-CoV-2 proven by DNA sequencing. These results were reported to the Connecticut Department of Public Health and published on July 17, 2020. Four days later on July 21, 2020, the Connecticut Department of Public Health also reported that a total of 90 out of 144 people tested between June 15 and July 17, many of whom were nursing home residents, received false-positive SARS-CoV-2 RT-qPCR test results because of a flaw in the test used at the state's public health laboratory. Gabrielle Fonrouge, *Connecticut lab finds 90 positive COVID-19 tests were false: report*, NEW YORK POST (Jul. 21, 2020),

https://nypost.com/2020/07/21/connecticut-testing-lab-botches-dozens-of-coronavirus-tests/. Attached as Exhibit 3 is a true and correct link to that article.

10. There is ample evidence that the currently used RT-qPCR tests are inaccurate for determining if a patient's sample in fact contains SARS-CoV-2, the virus causing COVID-19. The inherent flaw of PCR for the detection of SARS-CoV-2 RNA is further discussed as follows.

11. PCR (polymerase chain reaction) is a chemical reaction used to duplicate a defined segment of DNA exponentially in the test tube. To detect or to analyze a segment of target DNA molecule in question is usually by DNA sequencing, the process of determining the orders of the nucleotides (As, Ts, Cs, and Gs), which link up as a chain in the target DNA molecule.

12. However, the current technology cannot analyze one or a few DNA molecules in the sample being tested. These DNA molecules must be amplified, or made larger in number by a duplicating process to reach a mass of identical molecules to be analyzed. This amplification process, commonly referred to as PCR, is what Kary Mullis discovered, and consists of multiplying sequentially and exponentially by doubling the target DNA segment present in a test tube. So, 2 becomes 4, then becomes 8, then 16, and so forth, using newly formed copies as the

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 4 of 23

templates to make more new copies of the same molecule continuously. In such a duplication manner, each molecule of DNA in the original sample can become more than 1 billion copies after 30 cycles of amplification, in theory.

13. As noted, PCR multiplies DNA. But the genetic material that comprises the genome of SARS-CoV-2, the virus causing COVID-19, is RNA that is much more labile or unstable than DNA. It must be converted to DNA in order to utilize the PCR process. This is accomplished by action of an enzyme called reverse transcriptase (RT) in the first of four steps involved in the process. RT thus allows a single strand of RNA to be reverse-transcribed into a complementary strand of DNA, cDNA in short. The process of RT acting on RNA, leading to cDNA amplification through PCR, is called RT-PCR, which should be distinguished from RT-qPCR or rRT-PCR, the method currently being used for SARS-CoV-2 PCR testing.

14. The principle of PCR is based on primer-initiated and template-directed enzymatic polymerization of nucleotides. That means that there must be a segment of singlestranded DNA (ssDNA) serving as the template to direct the nucleotide incorporation for the synthesis of the new ssDNA whose sequence is complementary to the template. It also means that the synthesis of the new ssDNA must start with a primer, which is an oligonucleotide of about 20 nucleotides long and complementary to a segment of the target template DNA, annealing to (attached to) the target DNA at one of the two beginning sites of the target DNA to be amplified. Without a primer, the enzyme, DNA polymerase, will not work. In other words, PCR begins with enzymatic primer extension. The enzyme, a DNA polymerase, works like a type writer adding the nucleotides (A, C, G or T bases) one by one to the 3' end of the primer complementary to the sequence of the template in a 5' \rightarrow 3' direction.

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 5 of 23

15. The primer/template annealing process of PCR is based on DNA/DNA hybridization by forming hydrogen bonds between all complementary base pairs of the primer and template. However, DNA/DNA hybridization or annealing can take place even if there is only a partial match in base pairs between a primer and an unintended DNA in the PCR mixture in the absence of a fully matched target DNA template. Under certain circumstances the DNA polymerase can amplify an unintended (undesirable) DNA with a pair of partially matching primers and generate unintended (undesirable) PCR products. If the PCR products are not further analyzed for confirmation, false-positive test results may be produced.

16. The endpoint of an RT-qPCR test is arbitrarily set by the test kit manufacturer, using a Ct number to divide cases into "positive" or "negative." The extraordinary implications of this simple decision to frame results of the PCR testing as the basis for the entire "case numbers" tracking upon which virtually all public health measures are being based is almost incomprehensible.

17. I agree with Dr. Michael Mina, assistant professor of epidemiology at the Harvard T. H. Chan School of Public Health, who is quoted in the New York Post article as saying that this oversimplified interpretation of PCR as positive or negative is "irresponsible." This relates to the following discussion of the amplification process, sometimes also referred to as cycles. Dr. Mina is quoted in the Harvard Magazine (8/3/20) as saying that Current PCR testing detects virus genome-related materials "long after the infected person has stopped transmitting the virus." He further states "That means *the results are virtually useless for public health efforts to contain the raging epidemic.*" (emphasis added)

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 6 of 23

18. Tests with thresholds so high may detect not only live virus, but also simple genetic fragments, leftover from past infection that poses no risk for current exposure to others, or from other unrelated nucleic acids in the sample.

19. I agree with virologist Dr. Juliet Morrison that any test with a cycle threshold above 35 is too sensitive (in other words will read positive when the individual is not infectious).

20. I am of the opinion that it would be quite easy and simple to manipulate the number of positive results with this form of testing by the test kit manufacturers to please their customers whose business benefits from a high number of COVID-19 cases.

21. I believe that with varying numbers of cycles or amplifications being used in different states or even in different health systems in one state, it would be quite easy and simple to manipulate the number of positive results with this form of testing by simply changing the number of cycles to a higher number to produce the appearance of worsening or to a lower one to produce lower infection numbers. I agree with some experts who say that if over 40 amplifications be used, 100% of people tested might turn out to be positive.

22. I believe that the currently used RT-qPCR is a faulty diagnostic test, For example, an individual who gets a positive test result in a facility or area that is using a test setting the cutoff at 37 cycles might fly to another area where repeat testing is using a cutoff at 30 cycles and would likely get a negative test result. So, the same individual who "had COVID-19" in location one does not have it after flying to the second location.

23. This reveals the absurdity of the RT-qPCR based test.

24. To have children to take such an unreliable test is equally absurd.

I declare under penalty of perjury under the laws of the United States of America that the foregoing is true and correct.

Executed this 15th day of December, 2020 in Milford, Connecticut.

Signed

Sittanfler

Sin Hang Lee, MD

EXHIBIT 1

CURRICULUM VITAE

NAME:	SIN HANG LEE, M.D., F.R.C.P. (C), F.C.A.P.	
OFFICE ADDRESS:	Milford Molecular Diagnostics Laboratory 2044 Bridgeport Avenue, Milford, CT 06460	
PLACE OF BIRTH:	HONG KONG	
CITZENSHIP:	NATURALIZED U.S. CITIZEN 1976	
EDUCATION Tongji University Co Wuhan Medical Colle Hubei, China	llege of Medicine and ege (combined since 1952) Shanghai and Hankow,	1951-56
HIGH PROFESSIONAL DEGREE F.R.C.P. (C) Royal College of Physicians and Surgeons of Canada		
POSTGRADUATE TRAINI	NG and EXPERIENCE	
Teaching assistant in microbiology, Sichuan Medical College and Guiyang Medical College, China		
Demonstrator in path School of Medicine, I	ology, University of Hong Kong, Hong Kong	1961-63
Rotating clinical inter Baltimore, MD	rn, South Baltimore General Hospital,	1963-64
Resident, Assistant Pa Cornell Medical Cent	athologist and Pathologist at New York Hospital, er, New York, NY	1964-67
Pathology Fellow at N New York, NY	Memorial Hospital for Cancer and Allied Diseases,	1967-68
Assistant Professor of	f Pathology, McGill University, Montreal, Canada	1968-71
Associate Professor o	f Pathology, Yale University, New Haven, CT	1971-73

1973-2003
2004- 2015
2008-2015
2008- 2015 2015-

MEDICAL LICENSURE:

District of Columbia, New York and Connecticut (current), U.S.A. Licentiate of the Medical Council of Canada Certificate of full registration, General Medical Council, London, Great Britain

SPECIALTY BOARDS:

Diplomate, American Board of Pathology (AP)	1966
Certificated Specialist in General Pathology (AP and CP) Canada	1966

Expertise: General pathology, surgical pathology, clinical microbiology, and molecular diagnostics by PCR/direct DNA sequencing.

PUBLICATIONS:

- 1. Lee, S.H. Properdin (in Chinese) Chinese Med J. 8:796-799. 1958.
- Lee, S.H. Chinese translation of a handbook, "Die Praxis der Resistenz-und Spiegelbestimmungen zur Antibiotischen Therapie" by H-J. Otte and W. Köhler Veb Gustav Fischer Verlag-Jena. 1958, Peoples Hygiene Publisher, Beijing, China, 1961.
- 3. Lee, S.H. and Ts'O, T.O.T. Histological typing of lung cancers in Hong Kong. Brit. J. Cancer 17:37-40, 1963.
- 4. Lee, S.H. Histochemical demonstration of glutamic oxalacetic transaminase. Amer. J. Clin. Path. 49: 568-572. 1968.
- 5. Lee, S.H. and Torack, R.M. Aldehyde as fixative for histochemical study of glutamic oxalacetic transaminase. Histochem. 12: 341-344, 1968.
- 6. Lee, S.H. and Torack, R.M. The effects of lead and fixatives of glutamic oxalacetic transaminase. J. Histochem. 16: 181-184, 1968.
- 7. Lee, S.H. and Torack, R.M. Electron microscope studies of glutamic oxalacetic transaminase in rat liver cell. J. Cell Biol. 39: 716-724. 1968.

- 8. Lee, S.H. and Torack, R.M. A biochemical and histochemical study of glutamic oxalacetic transaminase activity of rat hepatic mitochondria fixed in situ and in vitro. J. Cell Biol. 39: 716-724. 1968.
- Lee, S.H. Ultrastructural localization of glutamic oxalacetic transaminase activity in cardiac muscle fiber and cardiac mitochondrial fraction of the rat. Histochem. 19: 99-109. 1969.
- 10. Lee, S.H. The possible role of the vesicles in renal ammonia excretion. J. Cell Biol. 45: 644-649. 1970.
- 11. Lee, S.H. Cytochemical study of in vivo inhibition of hepatic glutamic oxalacetic transaminase by hydrazine. Beitr. Path. 141: 99-106. 1970
- 12. Lee, S.H. and Aleyassine, H. Hydrazine toxicity in pregnant rats. Arch. Environ. Health 21: 615-619. 1970.
- 13. Chak, S.P. and Lee, S.H. Ultrastructural localization of glutamic oxaloacetic transaminase activity in adrenal cortical cell of rat. J. Ultrastructure Res. 35: 265-273. 1971.
- 14. Lee, S.H. and Aleyassine, H. Morphologic changes in the liver of mice bearing Ehrlich ascites tumor: Lab. Invest. 24: 513-522. 1971.
- 15. Aleyassine, H. and Lee, S.H. Inhibition by hydrazine, phenelzine and pargyline of insulin release from rat pancreas. Endocrinol. 89: 125-129. 1971
- Lee, S.H., Dusek, J. and Rona, G. Electron microscopic cytochemical study of glutamic oxalacetic transaminase activity in ischemic myocardium. J. Mol. Cell. Cardiol. 3: 103-109. 1971.
- 17. Aleyassine, H. and Lee, S.H. Inhibition of insulin release by substrates and inhibitors of monoamine oxidase. Amer. J. Physiol. 222: 565-569. 1972.
- 18. Lee, S.H. Isolation of parietal cells from glutaraldehyde-fixed rabbit stomach. J. Histochem. Cytochem. 20: 634-643. 1972.
- 19. Lee, S.H. Glutamate Oxalcetate Transaminase. In "Electron Microscopy of Enzymes" (M.A. Hayat, ed.). Vol. I. pp.116-130. Van Nostrand Reinhold Co., New York. 1973.
- 20. Lee, S.H. Ultracytochemistry of the mitochondrial glutamate oxalacetate transaminase activity. pp. 107-108. Proc. 4th Internatl. Congr. Histochem., Kyoto, Japan. 1972.
- Schachter, E.N., smith, G.J.W., Cohen, G.S., Lee, S.H., Lasser, A. and Gee, J.B.L. Pulmonary granulomas in a patient with pulmonary veno-occlusive disease. Chest 67: 487-489. 1975.

- 22. Lee, S.H. Estrogen binding of human breast cancer cells studied with a fluorescent estradiol conjugate. Fed. Proc. 37:462. 1978 (abstr.)
- 23. Lee, S.H. Cytochemical study of estrogen receptor in human breast cancer. Amer. J. Clin. Path. 70: 197-203. 1978.
- 24. Lee, S.H. Determination of breast cancer cell estrogen receptor in frozen sections. Lab. Invest. 40:268. 1979 (Abstr.)
- 25. Lee, S.H. Simultaneous detection of estrogen and progesterone receptors in breast cancer cells. Fed. Proc. 38:913. 1979 (Abstr.)
- 26. Lee, S.H. Cancer cell estrogen receptor of human mammary carcinoma. Cancer 44:1-12. 1979.
- 27. Lee, S.H. Cellular estrogen and progesterone receptors in mammary carcinoma. Amer. J. Clin. Path. 73:323-329. 1980.
- 28. Lee, S.H. Hydrophilic macromolecules of steroid derivatives for the detection of cancer cell receptors. Cancer 46:2825-2828. 1980.
- 29. Lee, S.H. Estrogen and progesterone receptors in breast cancer A new approach to measure. Connecticut Med. 44:622-625. 1980.
- Lee, S.H. Sex-steroid hormone receptors on mammary carcinoma. In Masson Monographs in Diagnostic Pathology. Diagnostic Immunohistochemistry. Ed. R. A. DeLellis. pp.149-164. Masson Publishing USA, Inc., New York.
- 31. Lee, S.H. The histochemistry of estrogen receptors. Histochem. 71:491-500. 1981.
- 32. Lee, S.H. Histochemical estrogen receptor assay. Amer. J. Clin. Path. 76:365. 1981
- Lee, S.H. Prospects for histochemical assay of steroid receptors In Endocrine Relationships in Breast Cancer. Ed. B. A. Stoll. pp.144-155. 1982 William Heinemann Medical Books LTD, London
- 34. Lee, S.H. Estrogen receptor-rich neuroglia of the rat brain. Lab. Invest. 46:49A 1982 (Abstr.)
- 35. Lee, S.H. Uterine epithelial and eosinophil estrogen receptors in rats during estrous cycle. Histochem. 74:443-452. 1982.
- 36. Lee, S.H. Estrogen-primed immature rat uterus a tissue control for histochemical receptor assay. Amer. J. Clin. Path. 79:484-486. 1983.

- 37. Lee, S.H. Augmentation and depletion of cytoplasmic estrogen-binding sites as visualized by histochemical technique. J. Steroid Biochem. 19:31S. 1983 (Abstr.)
- 38. Lee, S.H. Validity of a histochemical estrogen receptor assay. Supported by the observation of a cellular response to steroid manipulation. J. Histochem. Cytochem. 32:305-310. 1984.
- 39. Benz, C., Wiznitzer, I. and Lee, S.H. Flow cytometric analysis of fluorescein-conjugated estradiol (E-BSA-FITC) binding in breast cancer suspensions. Cytometry 6:260-267. 1985.
- Lee, S.H. Histochemical study of estrogen receptors in the rat uterus with a hydrophilic fluorescent estradiol conjugate. Localization of Putative Steroid Receptors. Vol. I. Eds. L.P. Pertschuk and S.H. Lee. CRC Press, Inc., Boca Raton, USA pp 59-83. 1985.
- 41. Lee, C., Jesik, J., Mangkornkanok, M., Sensibar, J. and Lee, S.H. Estrogen receptors and hormone responsiveness in serially transplanted mammary tumors in rats. Localization of putative Steroid Receptors. Vol. I Eds. L.P. Pertschuk and S.H. Lee. CRC Press, Inc. Boca Raton, USA, pp 85-93. 1985'
- Benz, C., Wiznitzer, I. and Lee, S.H. Flow cytometric analysis of fluorescent estrogen binding in cancer call suspensions. Localization of Putative Steroid Receptors. Vol. I Eds. L.P. Pertschuk and S.H. Lee, CRC Press, Inc. Boca Raton, USA. pp.95-110. 1985.
- 43. Lee, S.H. A fluorescent histochemical study of steroid receptors in human breast cancer. Localization of Putative Steroid Receptors. Vol. II Eds. L.P. Pertschuk and S.H. Lee. CRC Press, Inc. Boca Raton, USA pp. 37-50. 1985.
- 44. Lee, S.H., Charoenying, S., Brennan, T., Markowski, M. and Mayo, D.R. Comparative studies of three serologic methods for the measurement of *Mycoplasma pneumoniae* antibodies. Amer. J. Clin. Pathol. 92:342-347. 1989.
- 45. Lee, S.H. Coexistence of cytoplasmic and nuclear estrogen receptors. A histochemical study in human mammary cancer and rabbit uterus. Cancer 64:1461-1466. 1989.
- 46. Keefe, D.L., Michelson, D.S., Lee, S.H. AND Naftolin, F. Astrocytes within the hypothalamic arcuate nucleus contain estrogen-sensitive peroxidase, bind fluorescein-conjugated estradiol and may mediate synaptic plasticity in the rat.Amer. J. Obstet. Gynecol. 1991; 164:959-966.
- 47. Rao SK, Caride VJ;, Ponn R, Giakovis E, Lee H. F-18 fluorodeoxyglucose positron emission tomography-positive benign adrenal cortical adenoma: imaging features and pathologic correlation. Clinical nuclear medicine 2004;29:300-2.
- 48. Lee SH. Green tea consumption and mortality in Japan. JAMA 2007;297(4):360.
- 49. Lee SH. Expanded use of human papillomavirus testing in gynecologic practice. Amer J Clin Path. 2007;128(5):883-4.

- Lee, S. H., Vigliotti, V.S., Vigliotti, J.S. and Pappu, S. Routine human papillomavirus genotyping by DNA sequencing in community hospital laboratories. Infect Agent Cancer 2007; 2:11.
- 51. Lee, S. H., Vigliotti, V.S., and Pappu, S. DNA Sequencing Validation of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* Nucleic Acid Tests. Am J Clin Pathol. 2008;129:852-859.
- 52. Lee S.H., Vigliotti V.S., Pappu S. Human papillomavirus (HPV) infection among women in a representative rural and suburban population of the United States. Inter J Gyn Ob. 2009; 105:210-214.
- 53. Lee, S.H., Vigliotti, V.S., and Pappu, S. Molecular tests for human papillomavirus (HPV), Chlamydia trachomatis and Neisseria gonorrhoeae in liquid-based cytology specimen. BMC Women's Health 2009; 9:8.
- 54. Lee, S. H., Vigliotti, V.S., Vigliotti, J.S. and Pappu, S. Validation of human papillomavirus genotyping by signature DNA sequence analysis. BMC Clin Pathol 2009; 9:3.
- 55. Lee SH. HPV test is a virology test, not for predicting cancer. In Castle PE. "The evolving definition of carcinogenic human papillomavirus". Infect Agent Cancer 2009, 4:7.
- 56. Lee, S.H., Vigliotti, V.S., and Pappu, S. Signature sequence validation of human papillomavirus type 16 (HPV-16) in clinical specimens. J Clin Path. 2010;63:235-239.
- 57. Lee SH. HPV DNA test utilization. Am J Clin Path 2010;133(2):339.
- Lee, S. H., Vigliotti, V.S., Vigliotti, J.S., Jones W. and Pappu, S. Increased Sensitivity and Specificity of Borrelia burgdorferi 16S Ribosomal DNA Detection. Am J Clin Path. 2010; 133:569-576.
- 59. Lee SH. From human papillomavirus to cervical cancer. Obstet Gynecol 2010; 116:1221-1222.
- Lee, S. H., Vigliotti, V.S., Vigliotti, J.S., Jones, W., Williams, J., Walshon, J. Early Lyme disease with spirochetemia – diagnosed by DNA sequencing. BMC Res Notes. 2010 Nov 1; 3:273.
- 61. Lee SH, Castle PE, Stoler M, Kinney W. Patient Safety and the Next Generation of HPV DNA Tests. Am J Clin Pathol. 2011 Mar;135(3):481.
- 62. Lee SH: Chapter 5 in "Guidelines for the use of molecular tests for the detection and genotyping of human papilloma virus from clinical specimens." Methods Mol Biol 2012; 903:65-101.
- 63. Lee SH. Detection of human papillomavirus (HPV) L1 gene DNA possibly bound to particulate aluminum adjuvant in the HPV vaccine Gardasil®. J Inorg Biochem 2012; 117:85–92.

- 64. Lee SH. Detection of human papillomavirus L1 gene DNA fragments in postmortem blood and spleen after Gardasil® vaccination-A case report. Advances in Bioscience and Biotechnology 2012; 3: 1214-1224.
- 65. Lee SH. Topological conformational changes of human papillomavirus (HPV) DNA bound to an insoluble aluminum salt a study by low temperature PCR. Advances in Biological Chemistry 2013; 3: 76-85.
- 66. Hong G, Lee SH, Ge S, Zhou S. A Novel Low Temperature PCR Assured High-Fidelity DNA Amplification. International Journal of Molecular Sciences. 2013; 14:12853-12862.
- 67. Lee SH. Melting profiles may affect detection of residual HPV L1 gene DNA fragments in Gardasil®. Curr Med Chem. 2014; 21:932-940.
- 68. Lee SH, Vigliotti JS, Vigliotti VS, Jones W, Shearer DM. Detection of Borreliae in Archived Sera from Patients with Clinically Suspect Lyme Disease. International Journal of Molecular Sciences. 2014; 15:4284-4298.
- Lee SH, Vigliotti JS, Vigliotti VS, Jones W, Moorcroft TA, Lantsman K. DNA Sequencing Diagnosis of Off-Season Spirochetemia with Low Bacterial Density in Borrelia burgdorferi and Borrelia miyamotoi Infections. International Journal of Molecular Sciences. 2014; 15:11364-11386.
- 70. Lee SH, Vigliotti JS, Vigliotti VS, Jones W. From human papillomavirus (HPV) detection to cervical cancer prevention in clinical practice. Cancers (Basel). 2014; 6(4):2072-2099.
- 71. Lee SH, Zhou S, Zhou T, Hong G. Sanger Sequencing for BRCA1 c.68_69del, BRCA1 c.5266dup and BRCA2 c.5946del Mutation Screen on Pap Smear Cytology Samples. International Journal of Molecular Sciences. 2016; 17(2):229.
- 72. Lee SH. Lyme disease caused by Borrelia burgdorferi with two homeologous 16S rRNA genes: a case report. International Medical Case Reports Journal. 2016; 9:101–106.
- 73. Lambert JS, Cook MJ, Healy JE, Murtagh R, Avramovic G, Lee SH. Metagenomic 16S rRNA gene sequencing survey of Borrelia species in Irish samples of Ixodes ricinus ticks. PLoS One. 2019;14:e0209881.
- Lee SH, Healy JE, Lambert JS. Single Core Genome Sequencing for Detection of both Borrelia burgdorferi Sensu Lato and Relapsing Fever Borrelia Species. Int J Environ Res Public Health. 2019;16: E1779.
- 75. Lee S.H. Testing for SARS-CoV-2 in cellular components by routine nested RT-PCR followed by DNA sequencing. International Journal of Geriatrics and Rehabilitation. 2020; 2::69-96.

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 16 of 23

EXHIBIT 2

Sin Hang Lee, MD, F.R.C.P.(C) Milford Molecular Diagnostics Laboratory 2044 Bridgeport Avenue Milford, CT 06460 USA

March 22, 2020

Dr. Margaret Harris The World Health Organization's coronavirus response team harrism@who.int

Dr Eduardo Guerrero WHO Regional Office for the Americas <u>guerrere@paho.org</u>

Dr. Anthony S Fauci af10r@nih.gov

Extremely sensitive, no false-positive tests needed for SARS-CoV-2

Dear Drs. Harris, Guerrero and Fauci:

It has been widely reported in the social media that the RT-qPCR test kits used to detect SARS-CoV-2 RNA in human specimens are generating many false positive results and are not sensitive enough to detect some real positive cases, especially during convalescence.

RT-qPCR is known to generate false positive results when used to detect influenza A virus [1] and MERS-CoV, [2] another Coronavirus.

Without a nested (two-round) PCR, a single round RT-PCR may miss real infections caused by SARS-CoV [3] and by SARS-CoV-2 [4].

The major technical flaw of RT-qPCR for molecular diagnosis is the limitation of the length of its DNA probe which is about 25 bases long or shorter. And hybridization is not an accurate method to determine nucleotide sequences, the foundation of all nucleic acid-based diagnostics.

This letter recommends that the WHO coronavirus response team adopt or develop a nested RTqPCR protocol to generate a cDNA PCR amplicon to be used as the template for bi-directional sequencing. As demonstrated in this letter, nested RT-PCR is an extremely sensitive detection method and DNA sequencing will guarantee no-false positive results if all positive reports are accompanied by two-directional sequencing electropherograms, like an EKG for the diagnosis of Left Bundle Branch Block in a cardiologist's consultation.

Based on information retrieved from the GenBank databases and available in the public domain, there is a unique 398-base segment in the SARS-CoV-2 nucleocapsid (N) gene which not only has a 100% match with that in the Wuhan seafood market pneumonia virus, but also contains four single-nucleotide mutations found in the viruses isolated from patients in the states of

California, Texas and Massachusetts of the U.S.A. This segment of the gene can be targeted for accurate molecular diagnosis.

The nucleotide sequence of this 398-base gene segment is copied from the GenBank and reprinted here with the 4 mutated bases typed in red color. Identification of these virus isolates each with a single-base mutation in this segment may be useful in tracing the immediate source of the pathogen among patients and carriers tested positive for SARS-CoV-2.

Severe acute respiratory syndrome coronavirus 2 SARS-CoV-2 RNA Isolated from throat swab of patient in cruise ship, Japan, 02-10-2020 Sequence ID: <u>LC528233.1</u>

So	ore	Expect	Identities	Gaps	Strand	
/3	6 DILS(398)	0.0	398/398(100%)	0/398(0%)	Plus/Plus	
Quer	y 1	CAATCCTGCTAAC	AATGCTGCAATCGTGCTA	CAACTTCCTCAAGGAACA	ACATTGCCAAA 	60
Sbjc	t 28728	CAATCCTGCTAAC	AATGCTGCAATCGTGCTA	CAACTTCCTCAAGGAACA	ACATTGCCAAA	28787
Quer	ry 61	AGGCTTCTACGCA		AGTCAAGCCTCTTCTCGT'	ICCTCATCACG	120
Sbjc	t 28788	AGGCTTCTACGCA	GAAGGGAGCAGAGGCGGC	AGTCAAGCCTCTTCTCGT	TCCTCATCACG	28847
Quer	y 121	TAGTCGCAACAGT		GGCAGCAGTAGGGGAACT'	TCTCCTGCTAG	180
Sbjc	t 28848	TAGTCGCAACAGT	r <mark>c</mark> aagaaattcaactcca	GGCAGCA <mark>G</mark> TAGGGGAACT	TCTCCTGCTAG	28907
Quer	y 181	AATGGCTGGCAAT	GGCGGTGATGCTGCTCTT	GCTTTGCTGCTGCTTGAC	AGATTGAACCA	240
Sbjc	t 28908	AATGGCTGGCAAT	GGCGGTGATGCTGCTCTT	GCTTTGCTGCTGCTTGAC	AGATTGAACCA	28967
Quer	y 241	GCTTGAGAGCAAA	ATGTCTGGTAAAGGCCAA	CAACAACAAGGCCAAACT(GTCACTAAGAA	300
Sbjc	t 28968	GCTTGAGAGCAAA	ATGTCTGGTAAAGGCCAA	CAACAACAAGGCCAAACT	GTCACTAAGAA	29027
Quer	y 301	ATCTGCTGCTGAG	GCTTCTAAGAAGCCTCGG	CAAAAACGTACTGCCACT.	AAAGCATACAA	360
Sbjc	t 29028	ATCTGCTGCTGAG	GCTTCTAAGAAGCCTCGG	CAAAAACGTACTGCCACT	AAAGCATACAA	29087
Quer	y 361	TGTAACACAAGCT	ITCGGCAGACGTGGTCCA	GAACAAA 398		
Sbjc	t 29088	TGTAACACAAGCT	IT <mark>C</mark> GGCAGACGTGGTCCA	GAACAAA 29125		

NOTE: This 398-base sequence is identical to that of the Wuhan seafood market pneumonia virus, isolated in December 2019, GenBank Sequence ID: $NC_045512.2$

SARS CoV-2 isolates in the USA may have following single-base mutations in this segment at the positions typed in red (Sequences were retrieved from NCBI Databases).

29103 C>T Sputum of patient, TX, USA, 02-11-2020 Sequence ID: MT106054 28886 G>A Nasopharyngeal swab, CA, USA, 02-06-2020 Sequence ID: MT106052 28862 C>T Oropharyngeal swab, MA, USA, 01-29-2020 Sequence ID: MT039888 28792 A>T Nasopharyngeal swab, CA, USA, 01-23-2020 Sequence ID: MN994467



Left is an image of gel electrophoresis of the products of primary RT-PCR (upper half) and nested PCR (lower half) showing that nested PCR increases the sensitivity of RT-PCR at least 1,000 times in detecting SARS-CoV-2 RNA. The copy number of synthetic viral RNA added to each 25 µL primary RT-PCR mixture was calculated based on the analysis data supplied by BEI Resources, NIAID, NIH: Quantitative Synthetic RNA from SARS-Related Coronavirus 2, NR-52358. As demonstrated, this protocol can detect a single copy of viral RNA.

The 398-bp nested PCR amplicon shown in Lane 6 was used as the template for Sanger sequencing. The bi-directional sequences are pasted below.





Please inform your affiliated laboratories that we are now in position to assist them to resolve their questionable RT-qPCR test results with high Ct values (between 37 and 40) if they are able to send us 10 μ L of the residual RNA extract kept at -80°C in dry ice package. We will perform a nested RT-PCR on each of received residual samples, and perform a bi-directional Sanger sequencing on all positive cases and report the results back to the sender.

Contact person is: Sin Hang Lee, MD email shlee01@snet.net

Sincerely,

Hande

Sin Hang Lee, MD, F.R.C.P.(C)

References

- Martí NB, Del pozo ES, Casals AA, Garrote JI, Masferrer NM. False-positive results obtained by following a commonly used reverse transcription-PCR protocol for detection of influenza A virus. *J Clin Microbiol*. 2006;44(10):3845.
- 2. Pas SD, Patel P, Reusken C, et al. First international external quality assessment of molecular diagnostics for Mers-CoV. *J Clin Virol*. 2015;69:81–85.
- 3. Jiang SS, Chen TC, Yang JY, et al. Sensitive and quantitative detection of severe acute respiratory syndrome coronavirus infection by real-time nested polymerase chain reaction. Clin Infect Dis. 2004;38(2):293–296.
- 4. Nao, N., et al. Detection of second case of 2019-nCoV infection in Japan. 2020. https://www.who.int/docs/default-source/coronaviruse/method-niid-20200123-2.pdf?sfvrsn=fbf75320_7

Case 1:20-cv-09829-PGG Document 12-1 Filed 12/17/20 Page 21 of 23

EXHIBIT 3

NEWS

Connecticut lab finds 90 positive COVID-19 tests were false: report

By Gabrielle Fonrouge

July 21, 2020 | 1:38pm | Updated



Shutterstock

Sign up for our special edition newsletter to get a daily update on the coronavirus pandemic.

Nearly a hundred people in Connecticut who received positive coronavirus tests didn't actually have the disease, reports said.

A total of 90 out of 144 people tested between June 15 and July 17, many of whom are nursing home residents, received the false positives because of a flaw in the test used at the state's public health laboratory, NBC Connecticut reported.

State officials said the test manufacturer, the Food and Drug Administration and the Department of Public Health have all been notified about the snafu and immediate steps were taken to ensure patients were notified, the outlet reported.

"We have notified the healthcare facilities for everyone who received a false positive test result from our state laboratory," said Acting Commissioner Deidre S. Gifford from the state's department of social services.

"Accurate and timely testing for the novel coronavirus is one of the pillars supporting effective response to the COVID-19 pandemic."

Officials said the bad results came "from a widely-used laboratory testing platform that the state laboratory started using on June 15," the outlet reported.

Gifford said changes are already underway "to ensure the accuracy of future test results from this platform."

Moving forward, all positive coronavirus results will undergo further analysis from multiple scientists and if necessary, specimens will be retested using an alternate method, the outlet said.

Nursing home residents who received the false positive will be retested as soon as possible, the state's department of health said, according to the outlet.

FILED UNDER CONNECTICUT, CORONAVIRUS, LABORATORY, SAFETY TEST, 7/21/20